

14 TITLE

PROJECT NO.

BOOK NO.

Investigator: H. B. C. Date: 3/2/06
Notebook page #: 515/16 H Microsphere lot #: 90591-11Microsphere Production
Process Room ConditionsRoom Temp: 22°C
Room Humidity: 51.5% RH

Polymer Preparation

Polymer Type 1: PEG-120A
Source & Lot no.: BPTE Lot # 204-14-19
Mass (g): 3.60 g
Polymer type 2: D.L. PEA
Source & Lot no.: RE Lot # 31028
Mass (g): 3.61 g
Solvent Type: Water
Source & Lot no.: EM lot no 31068
Volume: 700 mL
Surfactant type: Levittol
Surfactant conc: 25 mg
Observation method: vis
Dissolution Temp: Room Temp
Dissolution Time: 15 min
Extraction 1: H₂O
Source & Lot no.: House DZ
Amount used: 200 mL

Comments:
Similar emulsion made in water as follows:
100 g in form of H₂O. Added to polymer solution.

Aeration Methodology

Sonication: None
Hom type: None
Frequency: None
Power: None
Temperature: None
Time: None
Time and spray: None
Spraying: None
Gas type: None
Gas pressure: None
Temperature: None
Time: None
Time and spray: None
Homogenization: NIETIS
Blade type: MACRO ultra-fine generator
Time: minutes (111)
Speed: 2.0 rpm (40)
Temperature: 20°C
Time and spray: 5 minutes
Comments:

Spray Conditions

Chemistry Proc.: Monomer solution, PEG-120A, PEA
Nozzle type: 0.7mm standard, vent D
Gas Pressure: 99 psi
Gas Flow rate: 600 L/H
Gas type: Medical grade
Feed Pressure: 112 psi
Inlet Temp: 22°C
Start Time: 1:16
End Time: 1:28
Mass Reproduc: 16.1-16.2 ± 0.1%
Yield (g): 1.7%

Process Conditions

| | Start 1:16 | End 1:27 | End 1:28 |
|----------------|------------|----------|----------|
| Outlet Temp: | 22°C | 22°C | 22°C |
| Filter Vacuum: | -40 mmHg | -10 mmHg | -10 mmHg |

Comments:
Did not get inlet tube as full down up 2 previous batches.

Drying Methodology

Type: Lyophilization, VIRTIS
Total dry time: 24 hours
Mass recovered: gasket (18,445-18,71) ± 1.7%
Yield (g): 1.805 g - 1.81 g - 2.5%
Comments:
1.4% in EtOH dry as before, dried from dry at 2.35
At 2.35 g, weight 1.81 g. Placed back in 1.81 g
Re-dried for 24 hours, 2.35 g.

Sizing Methodology

Room Temp: None
Room Humidity: None
Screening Time: None
Screen size: None
Screen type: None
Product recovered: None
Yield (g): None
Comments:

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H. B. C.

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John A. H. H.

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TJU Study For

PROJECT NO

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2nd

25 Samples weighed by Howard in Dry Box
on [redacted]. All but the 941082 and 941083
Samples sent to Forberg on [redacted] by Air mail
on dry ice / gel pack

EXECUTIVE UNDERPRODUCTIONS CHICAGO 60601 Made in U.S.A.

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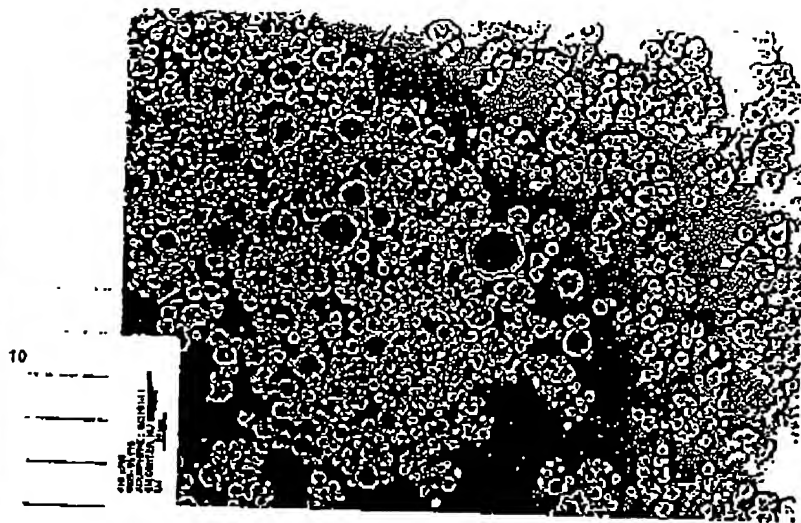
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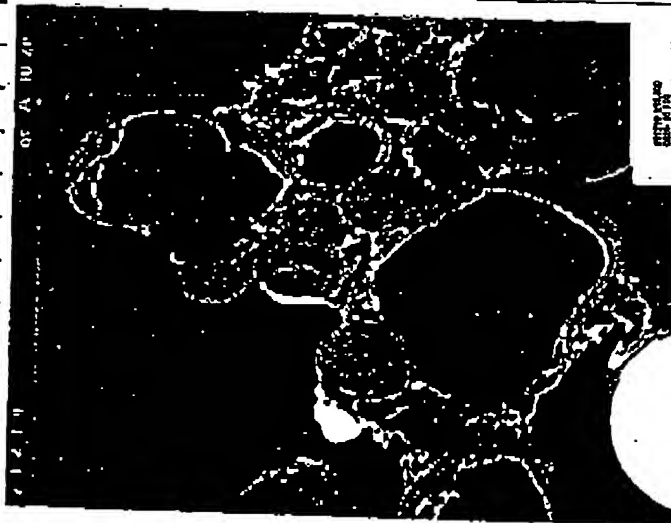


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Handwritten signature/initials

SCIENTIFIC IMAGERY PRODUCTIONS CHICAGO ILLINOIS Made in USA

SIGNATURE *James C. Shanks*
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DATE *[Redacted]* DATE *[Redacted]*

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PROJECT NO

VIA CONTINUED ...

Abstract

Notes

All samples will be prepared by weighing and modification.

All samples will be prepared in 30 ml. microcentrifuge vials. Empty 20 ml. microcentrifuge vials will be brought.

Saline (0.9%) will be used because of better flow characteristics in the pumping system. Prescribed sample of 100 g of NaCl will be brought out, and added to 1000 ml. with water in a bottle. Two more bottles will be kept as TPL. A total of 30 NaCl vials will be brought.

Vehicle 1 = 0.9% NaCl 20.7% glucose - reversed

Vehicle 2 = 0.9% NaCl 54.6 mg/ml. glucose

1) System eventually came as experiment, except 500ml saline in body

2) Vehicle 2 (VF) was used

3) Younger who did the entire study

4) After injection of sample, sample was shared, flow rate was then increased to 500-800ml/min until echogenic material detected by the oscilloscope, flow rate then dropped to 100-200 ml/min.

5) The later window moved dramatically with each pulse

6) Tubing was manipulated to remove bubbles. At least once (prior to injection 4) this resulted in change of alignment. At that one detection time, the machine was rechecked.

7) Cleaning procedure: (1) water pumped to remove all material, then saline pumped in (2) buffer emptied, saline added + pumped through.

(2) System pumped dry, saline pumped in

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400-30

| Injective | Sample ID/Amount | Exposition Preparation | Echogenicity Label | Echogenicity Over Time | Spun |
|-----------|------------------|------------------------|--------------------|------------------------|------|
| 1 | ALB-0.05 | NA (PABST) | not echogenic | | yes |
| 2 | ALB-0.50 | NA (PABST) | not echogenic | | yes |
| 3 | ALB-0.75 | NA (PABST) | not echogenic | | yes |
| 4 | ALB-1.00 | NA (PABST) | not echogenic | | yes |
| 5 | 3-VF-12ml | V/S1/V | not echogenic | | yes |
| 6 | 4-VF-12ml | V/S1/V | not echogenic | | yes |
| 7 | 5-VF-12ml | V/S1/V | not echogenic | | yes |
| 8 | 6-VF-12ml | V/S1/V | not echogenic | | yes |
| 9 | 7-VF-12ml | V/S1/V | not echogenic | | yes |
| 10 | 8-VF-12ml | V/S1/V | not echogenic | | yes |
| 11 | 9-VF-12ml | V/S1/V | not echogenic | | yes |
| 12 | 10-VF-12ml | V/S1/V | not echogenic | | yes |
| 13 | 11-VF-12ml | V/S1/V | not echogenic | | yes |
| 14 | 12-VF-12ml | V/S1/V | not echogenic | | yes |
| 15 | 13-VF-12ml | V/S1/V | not echogenic | | yes |
| 16 | 14-VF-12ml | V/S1/V | not echogenic | | yes |
| 17 | 15-VF-12ml | V/S1/V | not echogenic | | yes |
| 18 | 16-VF-12ml | V/S1/V | not echogenic | | yes |
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| 26 | 24-VF-12ml | V/S1/V | not echogenic | | yes |
| 27 | 25-VF-12ml | V/S1/V | not echogenic | | yes |
| 28 | 26-VF-12ml | V/S1/V | not echogenic | | yes |
| 29 | 27-VF-12ml | V/S1/V | not echogenic | | yes |
| 30 | 28-VF-12ml | V/S1/V | not echogenic | | yes |
| 31 | 29-VF-12ml | V/S1/V | not echogenic | | yes |
| 32 | 30-VF-12ml | V/S1/V | not echogenic | | yes |

400-30

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| 27 | 25-VF-12ml | V/S1/V | not echogenic | | yes |
| 28 | 26-VF-12ml | V/S1/V | not echogenic | | yes |
| 29 | 27-VF-12ml | V/S1/V | not echogenic | | yes |
| 30 | 28-VF-12ml | V/S1/V | not echogenic | | yes |
| 31 | 29-VF-12ml | V/S1/V | not echogenic | | yes |
| 32 | 30-VF-12ml | V/S1/V | not echogenic | | yes |

Shirley
Ruehl
TJW
on [redacted]

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